

SHENTEK

rcAAV-2/N Quantitation Kit

User Guide

Version: A/0

For Research Use Only

Product No.: 1403444-1

Reagents for 100 Reactions×2

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(IMPORTANT: Please read this document carefully before experiment.)

1. Product information

■ Product description

SHENTEK[®] rcAAV Quantitation Kit is suitable for qPCR detection of replication-competent adeno-associated virus (rcAAV) from cell culture harvested bulk and purified stock. This Kit is designed for the quantification of rcAAV-2/N contamination in serotypes of rAAV-2/N (N stands for possible different capsid serotypes). The sample types include but not limited to recombinant adeno-associated virus (rAAV) bulk and end-products, as well as harvested samples from cell culture desired for rcAAV detection.

The kit is for Research Use Only and is not for use in diagnostic procedures or for therapeutic purposes in humans or animals.

Key information before using this kit:

- AAV serotypes
- The inverted terminal repeat (ITR) sequence of the test sample rAAV need to match the following sequences:

> ITR sequence of rAAV-2/N

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TTGGCCACTCCCTCTCTGCGCGCTCGCTCGCTCACTGAGGCCGGGCGACC
AAAGGTCGCCCGACGCCCGGGCTTTGCCCGGGCGGCCTCAGTGAGCGAG
CGAGCGCGCAGAGAGGGAGTGGCCAACCTCCATCACTAGGGGTTTCCT
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■ Kit components and storage

WARNING: Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, mask, clothing and gloves.

Table 1. Kit components and storage

Reagent	Part No.	Quantity	Storage
T&R DNA Control-2	NNA027	lyophilized powder × 1 tube	-20°C
rcAAV qPCR Reaction Buffer	NNB009	400 µL × 4 tubes	-20°C, protect from light
Target Primer&Probe MIX-2	NNC048	200 µL × 1 tube	
Reference Primer&Probe MIX-2	NNC049	200 µL × 1 tube	
DNA Dilution Buffer (DDB)	NND001	1.5 mL × 4 tubes	-20°C
ddH ₂ O	NND010	1 mL × 1 tube	

Note: According to USP <659>, items with recommended storage temperatures not exceeding -20°C shall be stored within $\pm 10^{\circ}\text{C}$ (-30°C to -10°C). Short-term storage below -30°C is acceptable when supported by stability data.

The kit components can be stored under the appropriate conditions for up to 24 months. Please check the expiration date on the labels.

■ Applied instruments, including but not limited to the following

- SHENTEK-96S Real-Time PCR System
- ABI 7500 Real-Time PCR System
- CFX96 Real-Time PCR System
- LightCycler 480 II Real-Time PCR System

■ Required materials not included in the kit

- Low retention, DNase-free, Sterile microcentrifuge tubes, 1.5 mL
- Low retention, Sterile filter tips: 1000 μL , 100 μL and 10 μL
- 96-well qPCR plates with sealing film or PCR 8-strip PCR tubes with caps
- SHENTEK[®] Virus DNA & RNA Extraction Kit (Product No. 1506730)

■ Related equipments

- Benchtop microcentrifuge
- Real-Time PCR system
- Vortex mixer
- Pipettes: 1000 μL , 100 μL and 10 μL

■ Instructions for extraction and test

1. Samples start from harvested bulk

1.1 Extraction

- (1) Refer to the user guide of Virus DNA & RNA Extraction Kit for specific steps.
- (2) Benzonase Nuclease treatment of recombinant adeno-associated virus prior to extraction of the stock or final product is required to protect from interference of nucleic acid fragments that are not protected by viral capsid. Benzonase treatment

ensures more accurate results for the direct qPCR method.

1.2 Test

Depending on the gene being tested, different dilutions of purified sample solution are required:

- (1) When detecting the Reference gene, first dilute the purified sample solution into the range of standard curve ($20 - 2 \times 10^6$ copies/uL).
- (2) When detecting the Target gene, no dilution of the sample purification solution is required.

For example, if the concentration of rAAV virus stock solution is about 2×10^{12} vg/mL, then purified sample solution can be directly used for Target gene detection, but need to be diluted at least 1000 times before Reference gene detection.

2. Samples start from cultured cells

- (1) Titer of virus stock or final product can be quantified prior to infection of host cells, the sample can be extracted and purified according to the extraction procedure in 1.1, and use Reference gene sets to determine the concentration of rAAV;

Note: Dilute the extracted and purified solution of the test sample into the range of standard curve ($20 - 2 \times 10^6$ copies/uL).

- (2) After cell culture, the extraction process of test samples refer to the instruction manual of Virus DNA & RNA Extraction Kit for specific steps. The concentration of rcAAV was determined in using Target gene alone.

2. Methods

■ Experiment preparation

1. Irradiate the tabletop, pipettes and tubes with UV for 30 minutes, and disinfect with 75% ethanol.
2. Thaw the kit completely at $2-8^{\circ}\text{C}$ or melt on ice, vortex and spin briefly.

■ DNA Control serial dilutions for the standard curve

For the first use, Please check the concentration labeled on the tube containing the

T&R DNA Control-2 prior to dilution. Centrifuge the T&R DNA Control-2 at 12000 rpm for 1 min. Then pipette precisely 55 uL ddH₂O to the bottom of the tube to dissolve the lyophilized powder.

Note: To ensure that the lyophilized powder is fully dissolved, centrifuge briefly for 3-5 seconds, then gently invert to mix well, and repeat 3 times. If it stands for a long time, remix by repeating the above step once..

1. Thaw DDB completely at 2-8°C or melt on ice. Vortex to mix well and quickly spin down the reagents for 3-5 seconds in microcentrifuge, and repeat 3 times.
2. Label seven 1.5 mL microcentrifuge tubes: ST0, ST1, ST2, ST3, ST4, ST5 and ST6.
3. Dilute the DNA Control to 2×10^8 copies/uL with DDB in ST tube. Vortex to mix well and quickly spin down the ST tube for 3-5 seconds in microcentrifuge, and repeat 3 times to mix thoroughly.

Calculate the volume of DDB to prepare the ST:

$$\frac{\text{DNA Control conc. (A)} \times \text{Volume of DNA Control (B)}}{2 \times 10^8 \text{ copies}/\mu\text{L}} - \text{Volume of DNA Control (B)}$$

For example:

The concentration on the label of the DNA Control is 2.90×10^9 copies/ μL (A), pipette 10 μL (B) of the DNA Control to the ST tube. Add DDB in the volume shown in the calculation below to reach a final concentration of 2×10^8 copies/ μL .

$$\frac{2.90 \times 10^9 \text{ copies}/\mu\text{L} \times 10 \mu\text{L}}{2 \times 10^8 \text{ copies}/\mu\text{L}} - 10 \mu\text{L} = 135 \mu\text{L}$$

4. Add 90 μL DDB to ST0 and 180 μL DDB to each tube of ST1, ST2, ST3, ST4, ST5 and ST6.
5. Perform the serial dilutions according to Table 2:

Table 2. Dilution for T&R DNA Control

Serial dilution tube	Dilution	Conc. (copies/ μ L)
ST0	10 μ L ST + 90 μ L DDB	2×10^7
ST1	20 μ L ST0 + 180 μ L DDB	2×10^6
ST2	20 μ L ST1 + 180 μ L DDB	2×10^5
ST3	20 μ L ST2 + 180 μ L DDB	2×10^4
ST4	20 μ L ST3 + 180 μ L DDB	2×10^3
ST5	20 μ L ST4 + 180 μ L DDB	2×10^2
ST6	20 μ L ST5 + 180 μ L DDB	2×10^1

Note: The remaining unused DDB need to be stored at 2-8°C. If the solution is cloudy or contains precipitates, heat at 37°C until it clear.

Note: At least five concentration values should be included. when creating a standard curve. To select appropriate sample dilutions, we recommend to perform method validation before sample testing.

■ qPCR MIX preparation

- Determine the number of reaction wells based on your selected standard curve, with the number of test samples and control samples. Generally, triplicates are tested for each sample.
 - Number of Reference gene reaction wells = (6 standard points on the standard curve + 1 NTC + 1 NCS + 1 test samples) \times 3
 - Number of Target gene reaction wells = (6 standard points on the standard curve + 1 NTC + 1 NCS + 1 test samples) \times 3
- Prepare qPCR MIX referring to Table 3.

Table 3. qPCR MIX preparation

Reference gene		
Reagents	Volume/reaction	Volume for 30 reaction (includes 10% overage)
rcAAV qPCR Reaction Buffer	8 μ L	264 μ L
Reference Primer&Probe MIX-2	2 μ L	66 μ L
Total volume	10 μ L	330 μ L
Target gene		
Reagents	Volume/reaction	Volume for 30 reaction (includes 10% overage)
rcAAV qPCR Reaction Buffer	8 μ L	264 μ L
Target Primer&Probe MIX-2	2 μ L	66 μ L
Total volume	10 μ L	330 μ L

- Mix thoroughly and place on ice, aliquot 10 μ L/well into 96-well qPCR plate or PCR 8-strip tubes.

■ qPCR Reaction MIX preparation

- Prepare qPCR reactions following Table 4, and refer to a 96-well plate layouts template is shown in Table 5.

Table 4. qPCR Reaction MIX preparation

Standard curve	10 μ L qPCR MIX+20 μ L ST1 /ST2/ST3/ST4/ST5/ST6
NTC	10 μ L qPCR MIX+20 μ L DDB
NCS	10 μ L qPCR MIX+20 μ L purified NCS
Test sample	10 μ L qPCR MIX+20 μ L purified test sample

Table 5. Example of 96-well plate layouts

NCS Reference	NCS Reference	NCS Reference										A
NTC Reference	NTC Reference	NTC Reference				ST6 Reference	ST6 Reference	ST6 Reference	ST6 Target	ST6 Target	ST6 Target	B
1/x S- Reference	1/x S- Reference	1/x S- Reference				ST5 Reference	ST5 Reference	ST5 Reference	ST5 Target	ST5 Target	ST5 Target	C
						ST4 Reference	ST4 Reference	ST4 Reference	ST4 Target	ST4 Target	ST4 Target	D
						ST3 Reference	ST3 Reference	ST3 Reference	ST3 Target	ST3 Target	ST3 Target	E
NCS Target	NCS Target	NCS Target				ST2 Reference	ST2 Reference	ST2 Reference	ST2 Target	ST2 Target	ST2 Target	F
NTC Target	NTC Target	NTC Target				ST1 Reference	ST1 Reference	ST1 Reference	ST1 Target	ST1 Target	ST1 Target	G
S- Target	S- Target	S- Target										H
1	2	3	4	5	6	7	8	9	10	11	12	

Note: The example represents the detection of 6 concentration gradients of Reference gene standard curve (ST1-ST6) and Target gene standard curve (ST1-ST6), no template control NTC: NTC-Reference/NTC-Target, negative quality control NCS: NCS-Reference/NCS-Target, and diluted sample-Reference or Target, with 3 replicates for each sample.

Note: The plate layouts for sample loading can be adjusted based on the sample quantity.

2. For 1/x S-Reference wells, x refers to the number of dilution folds. The principle of dilution is that diluted test sample needs to be within the range of standard curve ($20-2 \times 10^6$ copies/uL).

■ qRT-PCR program setting

Note: The following instructions apply only to the ABI 7500 Real-Time PCR System with SDS v1.5.1. If you use a different instrument or software, refer to the applicable instrument or software documentation.

1. Create a new document, then in the Assay drop-down list, select Standard Curve (**Absolute Quantitation**).
2. In the Run Mode drop-down list, select **Standard 7500**, then click **Next**.

3. Click **New Detector**:
 - a. Enter Target-2 in the Name field, Select **FAM** in the Reporter Dye drop-down list and select **none** in the Quencher Dye drop-down list, then click **OK**
 - b. Enter Reference-2 in the Name field, Select **CY5** in the Reporter Dye drop-down list and select **none** in the Quencher Dye drop-down list, then click **OK**.
 - c. Select a color for the detector.
 - d. Select the detectors, then click **Add** to add the detectors to the document
4. Select **ROX** as the passive reference dye, then Click **Next**.
5. Select the applicable set of wells for the samples, then select Target-2 or Reference-2 detector for each well.
6. Select **Finish**, and then set thermal-cycling conditions:
 - a. Set the thermal cycling reaction volume to 30 μ L.
 - b. Set the temperature and time as following Table 6:

Table 6. qRT-PCR running program

Step	Temp.	Time(mm:sec)	Cycles
Pre-mutability	95°C	10 :00	1
Activation	95°C	00 :15	40
Denature	60°C	00 :30	
Anneal/extend	72°C*	00 :30**	

* Instrument will read the fluorescence signal during this step.


** For ABI 7500 instrument, please change this step from 72°C 30s to 72°C 34s, and keep 72°C 30s for other qPCR instruments.

■ Results analysis

1. Select **Set up** tab, then set tasks for each sample type by clicking on the Task Column drop-down list:
 - a. NTC: target DNA detector task = **NTC**
 - b. NCS, test samples: target DNA detector task = **Unknown**
2. Set up the standard curve as shown in the following table (Table 7):

Table 7. Settings for Standard curve

Tube label	Task	Quantity (copies/ μ L)
ST1	Standard	2×10^6
ST2	Standard	2×10^5
ST3	Standard	2×10^4
ST4	Standard	2×10^3
ST5	Standard	2×10^2
ST6	Standard	2×10^1

3. Select the **Results** tab, then select Amplification Plot.
4. In the Data drop-down list, select **Delta Rn vs Cycle**.
5. In the Analysis Settings window, enter the following settings:
 - a. Select **Manual Ct**.
 - b. In the Threshold field: Set the Threshold for the Reference-2 gene to 0.05; Set the Threshold for the Target-2 gene to 0.05.
 - c. Select **Automatic Baseline**.
6. Click the button  in the toolbar, then wait for the plate analysis to complete.
7. Select the **Result** tab > **Standard curve** tab, then verify the Slope, Intercept and R^2 values.
8. Select the Report tab, then achieve the mean quantity and standard deviation for each sample.
9. Select **File > > Export > > Results**. In the Save as type drop-down list, select **Results Export Files**, then click **Save**.
10. In the Report panel of Results, the 'Mean Quantity' column shows the detection values of NTC, NCS, test sample, in copies/ μ L.
The parameter settings of the result analysis should be based on the specific model and the software version, and generally can also be automatically interpreted by the instrument.
11. $Ct_{Reference}$ and Ct_{Target} values of NCS and NTC should be higher than the mean Ct value of the lowest concentration in the standard curve. If the validated quantitation limit (QL) concentration is lower than the lowest concentration in the standard curve, the value of the NCS should be below the QL.

12. The actual concentration of the test samples (copies/mL) need to be multiplied by its own dilution factors.
13. The contamination rate of rcAAV in rAAV is calculated according to the following equation:

Detection values of Target genes \div (Detection values of Reference genes \times dilution factor \div 2).

Note: Each rAAV contains two copies of Reference gene.

Effective date: 23 Apr. 2026

Support & Contact

The logo for SHENTEK, with the word in a bold, sans-serif font. The letters 'S', 'H', 'E', 'N', 'T', 'E', and 'K' are blue, while the letters 'S', 'H', 'E', and 'N' are green.

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